

How effective are antioxidants without fetal hemoglobin in tackling oxidative stress in sickle cell disease?

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Abstract

In our previous work titled "Antioxidative potential of foetal haemoglobin in sickle cell disease", we reported that fetal hemoglobin correlated directly with antioxidative markers and inversely with markers of oxidative stress in sickle cell subjects. Based on our observations, we concluded that fetal hemoglobin contributes to the cellular antioxidant defense system by limiting the rate of production of reactive oxygen species. In this current report, we aim to highlight the synergy between fetal hemoglobin and the antioxidant defense system in modulating oxidative stress in sickle cell disease.

Keywords: Sickle cell disease, Oxidative-stress, Fetal hemoglobin, Antioxidant defense mechanism, NRF2

Introduction

Sickle cell disorder (SCD) remains a disease of public interest, accounting for a significant number of deaths, especially in low and middle-income countries in Africa [1-3]. Despite years of intensive studies, its cure remains a subject of ongoing investigation [4,5]. The clinical manifestations of this disease including oxidative stress stem from a point mutation in the beta globin gene that produces sickle hemoglobin (HbS) [6-8]. Under anorexic conditions, HbS polymerizes to form the characteristic sickle shape structure of the erythrocytes in the sickle cell subjects [7,9]. The sickle red blood cells have a short life span which accounts for the high rate of hemolysis and anemia observed in sickle cell subjects [10,11]. These clinical events are further enhanced by vascular occlusion event, which partially or completely impedes the supply of nutrients required for cell survival [12,13].

Fetal Hemoglobin and Sickle Cell Disease

Fetal hemoglobin (HbF) is the predominant hemoglobin of a fetus, synthesized by a subset of red blood cells known as F-cells [14,15]. When compared to adult hemoglobin (HbA), HbF has a higher affinity for oxygen, making it more effective in transporting oxygen to the fetus in low oxygen environments such as the womb of a pregnant mother [16-18]. The structural variation between HbF ($\alpha_2\gamma_2$) and HbA ($\alpha_2\beta_2$) accounts for their different functions. Usually, the synthesis of HbF is gradually replaced with HbA a few months after delivery [19-21]. However, some subjects with SCD still retain the ability to synthesize a high concentration of HbF as adults. This genetic alteration has been reported to be beneficial, due to its ability to neutralize HbS polymerization: which is primarily responsible for all clinical complications associated with the disease.

HbF has been reported to affect the clinical manifestation of SCD [22,23]. While some subjects suffer a milder form of the disease, others are reported to suffer a more severe form of the disease defined by several episodes of crises. Several reports have associated subjects with a milder form

of the disease possessing higher HbF concentration in their blood cells compared to those with a low HbF in their red blood cells [24-26]. Based on these findings, it has been suggested that HbF is the strongest predictor of SCD severity.

Hemoglobin Switching

During human development, different forms of hemoglobin are typically produced by a process known as hemoglobin switching. At the early stage of gestation, embryonic hemoglobin ($\alpha_2\epsilon_2$) is the dominant hemoglobin expressed in the embryo. HbF ($\alpha_2\gamma_2$) expression soon takes over as the embryo develops into a fetus. After birth, the expression of HbF is gradually replaced by HbA ($\alpha_2\beta_2$) [27,28]. The physiological requirements of the various developmental stages are met by the expression of the various hemoglobin at their respective developmental stages [29,30]. The switching between the various hemoglobin types is genetically regulated by factors including genes, regulatory elements, and transcription factors.

Alteration in expression of transcription factors involved in the expression of HbF synthesis has been implicated in expression of abnormally high HbF concentration in some sickle cell subjects [31,32]. For instance, alteration in the expression of BCL11A, a transcription factor involved in the repression of γ -globin gene expression after birth, has been shown to cause the over-expression of γ -globin gene [33-35]. Similarly, an alteration in expression of the Kruppel-like factor-1 (KLF-1) transcription factor, involved in β -globin gene expression and γ -globin gene repression, has been reported to be responsible for the over-expression of HbF [36,37]. Hence, these molecules are being viewed as potential therapeutic targets for management of SCD due to their ability to alter HbF expression [38-41].

Oxidative Stress and Sickle Cell Disease

The pathophysiology of SCD is greatly influenced by oxidative stress [42-45], which occurs as a result of imbalance in the rate of production of reactive oxygen species (ROS) and antioxidant defense mechanism of the cell [46-48]. In this state, the synthesis of critical antioxidants required to neutralize the high amount of ROS generated is either compromised or depleted. Elevated production of ROS is responsible for clinical events including hemolysis, organ damage, and damage to deoxy-ribonucleic acid (DNA). In SCD, the generation of ROS is enhanced by sickle cell hemoglobin (HbS), which easily oxidizes to form superoxide anions, that are responsible for generating ROS. Furthermore, the product of hemolysis, including free hemoglobin and heme are responsible for the generation of ROS in SCD [49,50]. Due to variations in the rate of hemolysis among subjects with SCD, the rate of ROS generation and disease severity differs among subjects with the disorder. As a result, it has been suggested that HbF, which has the ability to neutralize the polymerization of HbS and reduce the rate of hemolysis, may function as an antioxidant [51].

Nuclear Factor Erythroid 2-related Factor-2 (NRF2): The Common Regulator Involved in Expression of Foetal Hemoglobin and Antioxidants

NRF2 plays a regulatory role in the expression of antioxidants in the body. Following oxidative stress, NRF2 stimulates the expression of antioxidants to clear the free radicals generated by clinical events such as hemolysis [52-55]. NRF2 achieves this aim either

by directly binding to the promoter region of genes encoding the different antioxidants to stimulate their expression or by enhancing the expression of other proteins involved in the metabolism of iron. The expression of iron-metabolizing proteins reduces the amount of iron needed for ROS synthesis [56,57]. NRF2 also regulates the expression of Glutathione; the most abundant antioxidant in the cell, by binding to and stimulating its expression [58,59]. Aside from the aforementioned roles of NRF2 in regulating the expression of antioxidants, it is also involved in the regulation of HbF expression.

NRF2 initiates the expression of HbF by binding to the promoter region of the γ -globin gene to stimulate its expression [60,61]. NRF2 also works alongside drug inducers such as simvastatin and hydroxy-urea to regulate the expression of HbF. The coordinated action of NRF2 and the drug inducers results in regulation of transcription factors which are directly involved in the expression of γ -globin gene [62,63]. For instance, studies have reported that the drugs, hydroxy-urea and simvastatin indirectly regulate the expression of the transcription factors BCL11A and KLF1, which are directly involve in regulating the expression of γ -globin gene by activating NRF2 transcription factor. Therefore, the action of NRF2 exacerbates oxidative stress in SCD by regulating the expression of antioxidants and HbF; the collective action of the two molecules greatly ameliorates the damaging effects of ROS in SCD.

Conclusion

The expression of high HbF in subjects with sickle cell disease is of immense benefit due to its ability to neutralize sickle cell hemoglobin polymerization and any other related clinical complications. The elevated rate of oxidative stress reported in subjects with low HbF concentrations compared to those with high HbF, suggests that antioxidant defense mechanism alone may be ineffective in tackling oxidative damage in SCD. Hence, a coordinated response of both antioxidants and HbF in sickle cell subject may be the best approach in tackling the elevated amount of ROS generated in SCD. Current studies are now focused on drugs that stimulate transcription factors such as NRF2, which is involved the regulation of expression of both HbF and antioxidants.

Conflict of Interest

Authors declare that no competing interests exist for any aspect of this commentary.

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References

1. Egesa WI, Nakalema G, Waibi WM, Turyasiima M, Amuje E, Kiconco G, et al. Sickle Cell Disease in Children and Adolescents: A Review of the Historical, Clinical, and Public Health Perspective of Sub-Saharan Africa and Beyond. International Journal of Pediatrics. 2022;2022(1):3885979.
2. Nwabuko OC, Onwuchekwa U, Iheji O. An overview of sickle cell disease from the socio-demographic triangle - a Nigerian single-institution retrospective study. Pan Afr Med J. 2022 Feb 23;41:161.
3. Williams TN. Sickle Cell Disease in Sub-Saharan Africa. Hematol Oncol Clin North Am. 2016 Apr;30(2):343-58.
4. Bell V, Varzakas T, Psaltopoulou T, Fernandes T. Sickle Cell Disease Update: New Treatments and Challenging Nutritional Interventions. Nutrients. 2024 Jan 15;16(2):258.

5. Salinas Cisneros G, Thein SL. Recent Advances in the Treatment of Sickle Cell Disease. *Front Physiol.* 2020 May 20;11:435.
6. Inusa BPD, Hsu LL, Kohli N, Patel A, Ominu-Evbota K, Anie KA, et al. Sickle Cell Disease-Genetics, Pathophysiology, Clinical Presentation and Treatment. *Int J Neonatal Screen.* 2019 May 7;5(2):20.
7. Elendu C, Amaechi DC, Alakwe-Ojimba CE, Elendu TC, Elendu RC, Ayabazu CP, Aina TO, Aborisade O, Adenikinju JS. Understanding Sickle cell disease: Causes, symptoms, and treatment options. *Medicine (Baltimore).* 2023 Sep 22;102(38):e35237.
8. Tebbi CK. Sickle Cell Disease, a Review. *Hemato.* 2022;3(2):341-66.
9. Rab MAE, van Oirschot BA, Bos J, Merckx TH, van Wesel ACW, Abdulmalik O, et al. Rapid and reproducible characterization of sickling during automated deoxygenation in sickle cell disease patients. *Am J Hematol.* 2019 May;94(5):575-84.
10. Akodu SO, Njokanma OF, AdeoluKehinde O. Erythrocyte indices in Pre-school Nigerian Children with Sickle Cell Anaemia in Steady State. *Int J Hematol Oncol Stem Cell Res.* 2015 Jan 1;9(1):5-9.
11. Darbari DS, Sheehan VA, Ballas SK. The vaso-occlusive pain crisis in sickle cell disease: definition, pathophysiology, and management. *European journal of haematology.* 2020 Sep;105(3):237-46.
12. King M, Walker L, Convery C, Davies E. Management of a Vascular Occlusion Associated with Cosmetic Injections. *J Clin Aesthet Dermatol.* 2020 Jan;13(1):E53-E58.
13. Adewoyin AS. Management of sickle cell disease: a review for physician education in Nigeria (sub-saharan Africa). *Anemia.* 2015;2015:791498.
14. Akinsheye I, Alsultan A, Solovieff N, Ngo D, Baldwin CT, Sebastiani P, Chui DH, Steinberg MH. Fetal hemoglobin in sickle cell anemia. *Blood.* 2011 Jul 7;118(1):19-27.
15. Steinberg MH. Fetal hemoglobin in sickle cell anemia. *Blood.* 2020 Nov 19;136(21):2392-400.
16. Pritišanac E, Urlesberger B, Schwabberger B, Pichler G. Fetal Hemoglobin and Tissue Oxygenation Measured With Near-Infrared Spectroscopy-A Systematic Qualitative Review. *Front Pediatr.* 2021 Aug 13;9:710465.
17. St. Jude Children's Research Hospital. (2022, October 12). A link between hypoxia and fetal hemoglobin provides hope for sickle cell disease. *ScienceDaily.* Retrieved on May 31, 2024.
18. Manning JM, Manning LR, Dumoulin A, Padovan JC, Chait B. Embryonic and Fetal Human Hemoglobins: Structures, Oxygen Binding, and Physiological Roles. In: Hoeger U, Harris J. (eds) *Vertebrate and Invertebrate Respiratory Proteins, Lipoproteins and other Body Fluid Proteins. Subcellular Biochemistry.* 2020;94. Springer, Cham. 6.
19. 3. Fetal hemoglobin: Structure and function. *Scandinavian Journal of Clinical and Laboratory Investigation.* 1982;42(sup160):32-7.
20. Simons M, Gretton S, Silkstone GGA, Rajagopal BS, Allen-Baume V, Syrett N, et al. Comparison of the oxidative reactivity of recombinant fetal and adult human hemoglobin: implications for the design of hemoglobin-based oxygen carriers. *Biosci Rep.* 2018 Jul 2;38(4):BSR20180370.
21. Paikari A, Sheehan VA. Fetal haemoglobin induction in sickle cell disease. *Br J Haematol.* 2018 Jan;180(2):189-200.
22. Antwi-Boasiako C, Frimpong E, Ababio GK, Dzudzor B, Ekem I, Gyan B, Sodji-Tettey NA, Antwi DA. Sickle Cell Disease: Reappraisal of the Role of Foetal Haemoglobin Levels in the Frequency of Vaso-Occlusive Crisis. *Ghana Med J.* 2015 Jun;49(2):102-6.
23. Mpalampa L, Ndugwa CM, Ddungu H, Idro R. Foetal haemoglobin and disease severity in sickle cell anaemia patients in Kampala Uganda. *BMC Blood Disorders.* 2012 Dec;12:11.
24. Adeodu OO, Akinlosotu MA, Adegoke SA, Oseni SBA. Foetal Haemoglobin and Disease Severity in Nigerian Children with Sickle Cell Anaemia. *Mediterr J Hematol Infect Dis.* 2017 Nov 1;9(1):e2017063.
25. Steinberg MH. Sickle cell anemia and fetal hemoglobin. *Am J Med Sci.* 1994 Nov;308(5):259-65.
26. Fitzhugh CD, Hsieh MM, Allen D, Coles WA, Seamon C, Ring M, Zhao X, Minniti CP, Rodgers GP, Schechter AN, Tisdale JF, Taylor JG 6th. Hydroxyurea-Increased Fetal Hemoglobin Is Associated with Less Organ Damage and Longer Survival in Adults with Sickle Cell Anemia. *PLoS One.* 2015 Nov 17;10(11):e0141706.
27. Sankaran VG, Xu J, Orkin SH. Advances in the understanding of haemoglobin switching. *Br J Haematol.* 2010 Apr;149(2):181-94.
28. Wang X, Thein SL. Switching from fetal to adult hemoglobin. *Nat Genet.* 2018 Apr;50(4):478-80.
29. Manning LR, Russell JE, Padovan JC, Chait BT, Popowicz A, Manning RS, et al. Human embryonic, fetal, and adult hemoglobins have different subunit interface strengths. Correlation with lifespan in the red cell. *Protein Sci.* 2007 Aug;16(8):1641-58.
30. Manning LR, Popowicz AM, Padovan J, Chait BT, Russell JE, Manning JM. Developmental expression of human hemoglobins mediated by maturation of their subunit interfaces. *Protein Sci.* 2010 Aug;19(8):1595-9.
31. Wilber A, Nienhuis AW, Persons DA. Transcriptional regulation of fetal to adult hemoglobin switching: new therapeutic opportunities. *Blood.* 2011 Apr 14;117(15):3945-53.
32. Mussolino C, Strouboulis J. Recent Approaches for Manipulating Globin Gene Expression in Treating Hemoglobinopathies. *Front Genome Ed.* 2021 Aug 2;3:618111.
33. Basak A, Sankaran VG. Regulation of the fetal hemoglobin silencing factor BCL11A. *Ann N Y Acad Sci.* 2016 Mar;1368(1):25-30.
34. Liu N, Xu S, Yao Q, Zhu Q, Kai Y, Hsu JY, et al. Transcription factor competition at the γ -globin promoters controls hemoglobin switching. *Nat Genet.* 2021 Apr;53(4):511-520.
35. Bauer DE, Orkin SH. Hemoglobin switching's surprise: the versatile transcription factor BCL11A is a master repressor of fetal hemoglobin. *Curr Opin Genet Dev.* 2015 Aug;33:62-70.
36. Caria CA, Faà V, Ristaldi MS. Krüppel-Like Factor 1: A Pivotal Gene Regulator in Erythropoiesis. *Cells.* 2022 Sep 29;11(19):3069.
37. Vinjamur DS, Alhashem YN, Mohamad SF, Amin P, Williams DC Jr, Lloyd JA. Krüppel-Like Transcription Factor KLF1 Is Required for Optimal γ - and β -Globin Expression in Human Fetal Erythroblasts. *PLoS One.* 2016 Feb 3;11(2):e0146802.
38. Wilber A, Nienhuis AW, Persons DA. Transcriptional regulation of fetal to adult hemoglobin switching: new therapeutic opportunities. *Blood.* 2011 Apr 14;117(15):3945-53.
39. Bauer DE, Kamran SC, Orkin SH. Reawakening fetal hemoglobin: prospects for new therapies for the β -globin disorders. *Blood.* 2012 Oct 11;120(15):2945-53.
40. Peralta R, Low A, Booten S, Zhou D, Kim A, Freier S, et al. Targeting KLF1 for the Treatment of Sickle Cell Disease Using Antisense Oligonucleotides. *Blood.* 2014 Dec 6;124(21):4038.
41. Starlard-Davenport A, Gu Q, Pace BS. Targeting Genetic Modifiers of

- HBG Gene Expression in Sickle Cell Disease: The miRNA Option. *Mol Diagn Ther.* 2022 Sep;26(5):497-509.
42. Vona R, Sposi NM, Mattia L, Gambardella L, Straface E, Pietraforte D. Sickle cell disease: role of oxidative stress and antioxidant therapy. *Antioxidants* (Basel). 2021 Feb 16;10(2):296.
43. Wang Q, Zennadi R. The Role of RBC Oxidative Stress in Sickle Cell Disease: From the Molecular Basis to Pathologic Implications. *Antioxidants* (Basel). 2021 Oct 13;10(10):1608.
44. Chirico EN, Pialoux V. Role of oxidative stress in the pathogenesis of sickle cell disease. *IUBMB Life.* 2012 Jan;64(1):72-80.
45. Silva DGH, Belini Junior E, de Almeida EA, Bonini-Domingos CR. Oxidative stress in sickle cell disease: an overview of erythrocyte redox metabolism and current antioxidant therapeutic strategies. *Free Radic Biol Med.* 2013 Dec;65:1101-9.
46. Burton GJ, Jauniaux E. Oxidative stress. *Best practice & research Clinical obstetrics & gynaecology.* 2011 Jun 1;25(3):287-99.
47. Pizzino G, Irrera N, Cucinotta M, Pallio G, Mannino F, Arcoraci V, Squadrito F, Altavilla D, Bitto A. Oxidative Stress: Harms and Benefits for Human Health. *Oxid Med Cell Longev.* 2017;2017:8416763.
48. Birben E, Sahiner UM, Sackesen C, Erzurum S, Kalayci O. Oxidative stress and antioxidant defense. *World Allergy Organ J.* 2012 Jan;5(1):9-19.
49. Gbotosho OT, Kapetanaki MG, Kato GJ. The Worst Things in Life are Free: The Role of Free Heme in Sickle Cell Disease. *Front Immunol.* 2021 Jan 27;11:561917.
50. Wang Q, Zennadi R. The Role of RBC Oxidative Stress in Sickle Cell Disease: From the Molecular Basis to Pathologic Implications. *Antioxidants* (Basel). 2021 Oct 13;10(10):1608.
51. Dogonzo YI, Onyeabor CC, Oru CM, Owusi OD, Ozor RC, Ebubechi O, et al. Antioxidative Potential of Foetal Haemoglobin in Sickle Cell Disease. *International Journal of Research and Reports in Hematology.* 2023 Sep 6;6(2):179-84.
52. Ngo V, Duennwald ML. Nrf2 and Oxidative Stress: A General Overview of Mechanisms and Implications in Human Disease. *Antioxidants* (Basel). 2022 Nov 27;11(12):2345.
53. Ma Q. Role of nrf2 in oxidative stress and toxicity. *Annu Rev Pharmacol Toxicol.* 2013;53:401-26.
54. Boas SM, Joyce KL, Cowell RM. The NRF2-Dependent Transcriptional Regulation of Antioxidant Defense Pathways: Relevance for Cell Type-Specific Vulnerability to Neurodegeneration and Therapeutic Intervention. *Antioxidants* (Basel). 2021 Dec 21;11(1):8.
55. Tonelli C, Chio IIC, Tuveson DA. Transcriptional Regulation by Nrf2. *Antioxid Redox Signal.* 2018 Dec 10;29(17):1727-45.
56. Vogt AS, Arsiwala T, Mohsen M, Vogel M, Manolova V, Bachmann MF. On Iron Metabolism and Its Regulation. *Int J Mol Sci.* 2021 Apr 27;22(9):4591.
57. Mariani R, Trombini P, Pozzi M, Piperno A. Iron metabolism in thalassemia and sickle cell disease. *Mediterr J Hematol Infect Dis.* 2009 Oct 27;1(1):e2009006.
58. Bell KF, Fowler JH, Al-Mubarak B, Horsburgh K, Hardingham GE. Activation of Nrf2-regulated glutathione pathway genes by ischemic preconditioning. *Oxid Med Cell Longev.* 2011;2011:689524.
59. Harvey CJ, Thimmulappa RK, Singh A, Blake DJ, Ling G, Wakabayashi N, et al. Nrf2-regulated glutathione recycling independent of biosynthesis is critical for cell survival during oxidative stress. *Free Radic Biol Med.* 2009 Feb 15;46(4):443-53.
60. Zhu X, Oseghale AR, Nicole LH, Li B, Pace BS. Mechanisms of NRF2 activation to mediate fetal hemoglobin induction and protection against oxidative stress in sickle cell disease. *Exp Biol Med* (Maywood). 2019 Feb;244(2):171-82.
61. Zhu X, Li B, Pace BS. NRF2 mediates γ -globin gene regulation and fetal hemoglobin induction in human erythroid progenitors. *Haematologica.* 2017 Aug;102(8):e285-e288.
62. Xi C, Palani C, Takezaki M, Shi H, Horuzsko A, Pace BS, Zhu X. Simvastatin-Mediated Nrf2 Activation Induces Fetal Hemoglobin and Antioxidant Enzyme Expression to Ameliorate the Phenotype of Sickle Cell Disease. *Antioxidants* (Basel). 2024 Mar 11;13(3):337.
63. Macari ER, Schaeffer EK, West RJ, Lowrey CH. Simvastatin and t-butylhydroquinone suppress KLF1 and BCL11A gene expression and additively increase fetal hemoglobin in primary human erythroid cells. *Blood.* 2013 Jan 31;121(5):830-9.